

TISSUE CULTURE PROCEDURE



STEP 4: TRANSFER

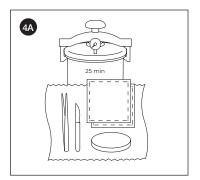
SCAN to watch our Tissue Culture Procedure Video:

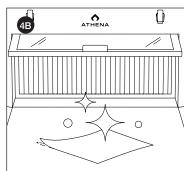


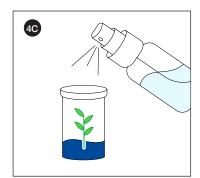
⚠ DISCLAIMER:

- All sterile work is to be done under the workzone unless stated otherwise.
- Always wear gloves and a face covering when working in the workzone.
- Any container with liquid placed in the Autoclave must have a loosely opened lid.
- Spray gloves with alcohol between processes in the workzone.
- **4A.** Wrap the **forceps**, **scalpel**, **work surface**, and **paper towel** in aluminum foil to place in the **Autoclave** for a full cycle.
- 4B. Clean and sterilize the Flow Hood workzone with alcohol wipes (inside and front surfaces).
- **4C.** Use the alcohol sprayer to sterilize all **culture vessels** filled with **Roots/ Shoots** formula, the vessels with explants, and the **alcohol burner** before placing them into the **workzone**.
- 4D. Place the items from the Autoclave directly under the Flow Hood.
- 4E. Remove an explant showing prominent new growth from the Shoots culture vessels and place it on the work surface.
 - Note: Ensure that the tools do not touch the media or culture vessels.

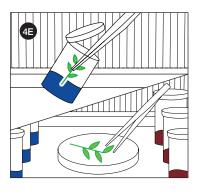
- **4F.** Dissect the explant at the middle of each internode leaving enough stem under each node to go into the media.
- 46. If the goal is to preserve the genetic material: Place the cutting into a culture vessel with Shoots media.
- **4H.** If the goal is to create a mother plant: Place the cutting into a **culture vessel** with Roots media.
- 41. Sterilize the forceps and scalpel blade with the alcohol burner after each cutting.
- 4J. Spray the work surface with alcohol after each cutting. Note: Repeat steps 4E-4J for each explant dissected.
- **4K.** When all **culture vessels** are prepared with newly made explants, store them under a clone light at 75-125 PPFD and 20-25.5°C (68-78°F) room temperature.

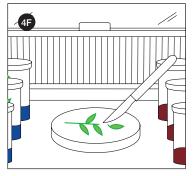


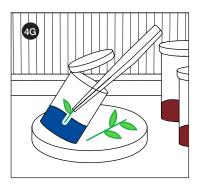


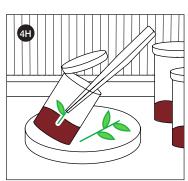


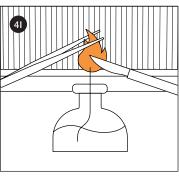


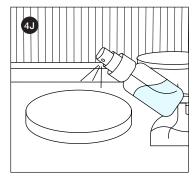




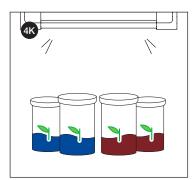








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